

Overview of Facility Software for Biomolecular NMR

by

Chi-Fon Chang

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National Program for Genomic Medicine High Field NMR Core Facility,
The Genomic Research Center, Academia Sinica

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Facility software for biomolecular NMR

NMR data processing software

•XWINNMR (process NMR data on IRIX 6.X & Linux)

License required

•nmrPipe (process NMR data on IRIX6.X & Linux)

NMR data analysis software

•AURELIA (analyze NMR data on IRIX 6.X & Linux)

License required

•nmrDraw (analyze NMR data on IRIX 6.X & Linux)

•nmrView (analyze NMR data on IRIX 6.X & Linux)

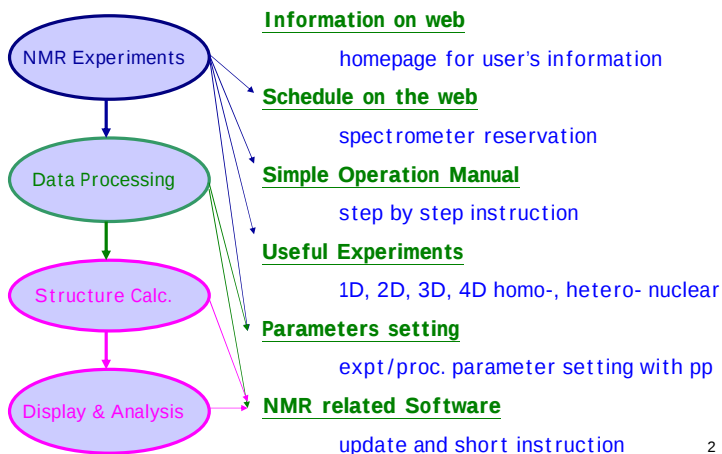
•Sparky (analyze NMR data on Linux)

NMR auto assignment

•TATAPRO (on IRIX)

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NMR Core Facility Service



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Structure Calculation related software

•CSI Chemical Shift Index (making consensus plot on IRIX)

•TALOS (dihedral angles prediction on IRIX 6.X & Linux)

•XPLOR (structure calculation on IRIX)

•CNS (structure calculation on IRIX6.X & Linux)

•ARIA (auto NOE assign and structure calculation on IRIX6.X & Linux)

•CYANA (auto NOE assign and structure calculation on IRIX6.X & Linux)

License required

Structure display & analysis software

•PROCHECK (structure analysis on IRIX)

•INSIGHT II (on IRIX , license from computer center)

•MOLMOL (on IRIX & Linux)

•GRASP (on IRIX) *License required*

NMR dynamics

•MODELFREE (on IRIX, Linux & Euler)

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Tips for using facility software

XWINNMR (process NMR data)

- Input data : data acquired from xwinnmr
- How to run:
`> xwinnmr` (or , for PC, click on xwinnmr icon)

AURELIA (analyze NMR data)

- Input data: data processed by xwinnmr (2rr, 3rrr, ...)
- How to run:
`> aurelia filename`

nmrView (analyze NMR data) <http://www.nmrview.com/>

- Input data: data processed by nmripe or felix
- How to run:
`> nmrvview`

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CSI (making consensus plot) <http://www.pence.ualberta.ca/ftp/csi/csi.html/>

- Input data:
 1. chemical shift table for HA, CA, CB, CO

#	AA	HA	CA	CB	CO
1	A	4.15	51.8	18.8	174.0
 2. average value for Gly HA, "0" for unassigned, "C" for oxidized cysteine , "B" for reduced cysteine
- How to run: (the same directory as input data)
`> csi` (the following the instruction)
- Output data:
 1. text output
 2. graphical output

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TATAPRO (auto-assignment)

- Input data:
 1. peak picking for HNCBCA, CBCA(CO)NH, HNCO, HNCACO
 2. sequence in one letter code
 3. pml.inp
 4. assign.inp
- How to run: (the same directory as input data)
`> pml` (to create "master_list"
`> assign` (start assignment)
- Output data:
 1. master_list (from pml)
 2. assignment.final (final assignment list)

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TALOS (dihedral angles prediction) <http://spin.niddk.nih.gov/bax/software/TALOS/>

- Input data: chemical shift table for N, HA, CA, CB, CO

1	M	HA	4.15
1	M	C	170.54
1	M	CA	54.82
1	M	CB	33.27
2	Q	N	123.01
- How to run: (the same directory as input data)
`> source nmripe`
`> talos.tcl -in myshift.tab` (to perform the database searches)
`> vina.tcl -in myshift.tab -auto` (to summarized the results)
`> vina.tcl -in myshift.tab -ref mystruct.pdb -auto`
`> rama.tcl -in myshift.tab` (to inspect and adjust the predictions)
- Output data:
 pred.tab with predicted phi, psi value

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ARIA (auto NOE assign and structure calculation)

<http://www.pasteur.fr/recherche/unites/Binfs/aria/>

- Input data:
 1. complete chemical shift table and 3 letter sequence file
 2. peak picking table for NOESY type experiment
 3. additional H-bond, CSI, RDC, or dihedral angles constraint
- How to run:
 - > *source aria*
 - > *netscape aria.html* (to generate new.html file)
 - > *aria* (to generate a "working directory" , and run.cns)
 - > *netscape aria.html* (to edit parameter for structure cal.)
 - > *aria* (start structure calculation based on run.cns)
- Output data:
 - Structures in pdb
 - NOE assignment table
 - Structures analysis using prochecknmr

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INSIGHT II (structure display, license from computer center)

- Input data : pdb
- How to run:
 - > *source insightII*
 - > *insightII*

MOLMOL (structure display) <http://www.mol.biol.ethz.ch/wuthrich/software/molmol/>

- Input data : pdb
- How to run:
 - > *molmol*

GRASP (structure display) <http://honiglab.cpmc.columbia.edu/grasp/>

- Input data : pdb
- How to run:
 - > *grasp*

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CYANA (auto NOE assign and structure calculation)

•Input data :

1. complete chemical shift table and 3 letter sequence file
 2. peak picking table for NOESY type experiment
 3. additional H-bond, CSI, RDC, or dihedral angles constraint
 4. init.cya
 5. CANDID.cya
- How to run:
 - > *cyana*
 - > *cyana> CANDID*
 - Output data: (ex: Ramachandran plot)

PROCHECK (structure analysis)

http://www.biochem.ucl.ac.uk/~roman/procheck_nmr/procheck_nmr.html

- Input data : summary of all structures in pdb
- How to run:
 - > *procheck*
 - > *procheck_nmr inputname*
- Output data: (ex: Ramachandran plot)

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MODELFREE (NMR Dynamic Data Analysis)

<http://cpmcnet.columbia.edu/dept/qsas/biochem/labs/palmer/software/modelfree.html>

- Input data:
 1. pdb file
 2. R1, R2, NOE data
 3. R2/R1 and d(R2/R1) data
- Analysis steps:
 - step1: PDBINERTIA* (input original pdb to obtain int.pdb)
 - step2: R2R1_TM* (input R2/R1 data to obtain local tm for each spin)
 - step3: QUADRIC_DIFFUSIOM*
(input step1&2 result to obtain Diso, Dration, Theta, Phi)
 - step4: ModelFree analysis*
(using data from above to do modelfree analysis)

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